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Role of mineral nutrition in minimizing cadmium accumulation by plants

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Abstract

Cadmium (Cd) is a highly toxic heavy metal for both plants and animals. The presence of Cd in agricultural soils is of great concern regarding its entry into the food chain. Cadmium enters into the soil – plant environment mainly through anthropogenic activities. Compounds of Cd are more soluble than other heavy metals, so it is more available and readily taken up by plants and accumulates in different edible plant parts through which it enters the food chain. A number of approaches are being used to minimize the entry of Cd into the food chain. Proper plant nutrition is one of the good strategies to alleviate the damaging effects of Cd on plants and to avoid its entry into the food chain. Plant nutrients play a very important role in developing plant tolerance to Cd toxicity and thus, low Cd accumulation in different plant parts. In this report, the role of some macronutrients (nitrogen, phosphorus, sulfur and calcium), micronutrients (zinc, iron and manganese), and silicon (a beneficial nutrient) has been discussed in detail as to how these nutrients play their role in decreasing Cd uptake and accumulation in crop plants.

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Keywords: accumulation; cadmium; edible parts; macronutrients; micronutrients; silicon

INTRODUCTION

Among all the heavy metals, cadmium (Cd) is considered to have high toxicity to humans and all other living organisms as it has no known biological functions in aquatic or terrestrial organisms.¹ Recent advances in industry and agriculture have led to an increased level of Cd in the agricultural soil environment. Cadmium enters the soil through various anthropogenic sources including application of phosphate fertilizers, waste water, Cd-contaminated sewage sludge and manures, and anthropogenic emissions from power stations, metal industries, urban traffic and cement industries.^{2–5} Due to its high mobility in soils, Cd accumulation in plants grown on Cd-contaminated soils poses a serious threat to human and animal health.⁶ High concentrations of Cd in rice and drinking water play havoc in *Itai-Itai* patients by lowering glomerular rates.^{7,8}

Cadmium enters the human body from many sources, i.e. soil, water, air, plants and animals, but the main source of Cd in non-occupational people except smokers is the consumption of Cd-contaminated food. Although it affects a number of organs in the human body, Cd mainly accumulates in the human kidney with a biological half-life of about 10–30 years (depending on the age), and causes renal tubular damage and pulmonary emphysema. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. And thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone. Salso disturbed by Cd, and thus causes hypercalciuria and the formation of kidney stone.

Ingestion of crops containing high amounts of Cd may contribute substantial Cd to the human diet.¹⁶ Of primary concern is

its transfer from vegetables to the body, because vegetables contribute $\geq 70\%$ of Cd intake in humans. 11 The joint FAO/WHO Expert Committee on Food Additives (JECFA), and the Codex Committee on Food Additives and Contaminants (CCFAC) have proposed a limit of 0.1 mg kg $^{-1}$ for Cd in cereals, pulses and legumes due to the risk associated with the long-term consumption of Cd-contaminated crops; 17,18 while the European Community has a limit of 0.2 mg kg $^{-1}$ for wheat grain, 19 and the maximum tolerable limit of Cd for humans proposed by FAO/WHO is 70 μg a day. 11,16

Through its effects on various biochemical and physiological processes in plants, Cd could inhibit plant growth and cause cell death above critical levels.^{20,21} Cadmium-induced growth reduction might be explained on the basis of inhibition of carbon fixation due to a decrease in photosynthetic rate and chlorophyll content.²² Cadmium in soils could induce water stress in plants by decreasing stomatal conductance, transpiration

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rate and leaf relative water contents.²³ This is the result of physiological damage from decreased intracellular space and amount of chloroplast, and cell enlargement.²⁴ Cadmium toxicity induces the production of reactive oxygen species (ROS) which may result in membrane damage, and the destruction of cellular organelles and biomolecules.²⁵ These damages can enhance the translocation of Cd to aerial parts. Moreover, the lack of energy for Cd sequestration in roots may also be responsible for its translocation from roots to aerial parts.

According to Verkleij et al.²⁶ there is a duality in plant tolerance to heavy metals. Certain heavy metals like iron (Fe), copper (Cu), zinc (Zn) and manganese (Mn) at low concentration are beneficial for plants either through improving plant growth, biofortification, or both, and in this way beneficial for all living organisms in the food chain. However, heavy metals also exert damaging effects on plants at similar or higher concentration depending on the specific/tolerance mechanisms the plants use to protect themselves, but could also produces compounds with some deleterious potential for organisms, including man. High levels of heavy metals in soils, including Cd, could damage biomemebranes and cause uncontrolled uptake or translocation of Cd in plants. In addition to this toxicity-induced Cd accumulation, plants may accumulate significant amounts of Cd in different parts without any significant toxicity and yield loss.²⁷ Hence, programs for safe food production from Cd-contaminated soils should concentrate on the minimization of Cd accumulation instead of the mitigation of Cd toxicity. Moreover, accumulation of Cd in edible plant parts is more important than total plant uptake.²⁸

It is necessary to minimize the entry of Cd into the food chain because of the number of associated health risks. Many strategies have been devised to minimize Cd toxicity and to check its entry into the food chain. Selection and breeding of crop plants/cultivars that accumulate low Cd in the grain and other edible plant parts is one of these approaches.^{29–32} It seems an attractive approach to change the Cd profile of crop plants and the benefit continues generation after generation in plants through the seed. Lower Cd durum wheat and sunflower cultivars have been introduced in the last few years; improving cultivars takes time and money, but provide the persistent benefit of lower dietary Cd. Differential Cd uptake and distribution may be the result of differences among cultivars in the capacity to retain absorbed Cd in the roots or to variations in xylem loading or Cd retranslocation in the phloem. 33,34 Structural and physiological characteristics of cultivars may lead to different capabilities of retaining Cd inside the roots and, in turn, to the observed variation in Cd partitioning.³³ However, there are constraints to using this approach to produce low-Cd food as it is very time consuming to develop and test a new cultivar. Generally, it takes 5–10 years to develop new cultivars. 35 Moreover, low Cd accumulation adds an additional trait to an already existing long list (yield, quality, disease resistance and many other characteristics) required of a new cultivar.³⁵ It also implies that it is very difficult to manage the low Cd accumulation trait with a number of other characteristics in the same cultivar.

Phytoextraction is another approach to minimize Cd entry into the food chain, which involves the use of hyper-accumulator plants to remove Cd from soil. 36,37 But the problem is that hyper-accumulator plants are slow growing and produce very low biomass and a long time is required, perhaps several years, to remediate the contaminated site. Moreover, the disposal of Cd-accumulated plant material is also of great concern and methods for the disposal of metal-enriched biomass have not yet been well developed. Similarly, crop rotation, liming and the use

of other organic and inorganic amendments⁴⁰ are some other approaches being used to remediate Cd-contaminated soils, but these approaches are time consuming and require extra resources.

Use of plant nutrients to alleviate Cd toxicity in plants is a relatively inexpensive, time saving and effective approach to avoid Cd contamination of food. Additionally, for better plant growth an adequate and balanced supply of nutrients is essential in proper amounts and at the correct time. Growers are already applying nutrients to obtain good crop yield, so to alleviate Cd toxicity the proper management of these plant nutrients is needed, keeping in mind the interactions between Cd and plant nutrients. Several of the plant nutrients have many direct as well as indirect effects on Cd availability and toxicity. Direct effects include decreased Cd solubility in soil by favoring precipitation and adsorption, 41-44 competition between Cd and plant nutrients for the same membrane transporters, 45-47 and Cd sequestration in the vegetative parts to avoid its accumulation in grain/edible parts.^{48–50} Indirect effects include dilution of Cd concentration by increasing plant biomass⁵¹ and alleviation of physiological stress.52-56 Relatively high doses of N and some other plant nutrients can cause soil acidification,⁵⁷ enhancing the solubility and bioavailability of heavy metals including Cd.50 Moreover, fertilizers determine Cd speciation and complex formation, which could affect Cd movement to roots and its absorption into the roots. Fertilizer application also affects the rhizosphere composition, root growth and overall plant growth,⁵⁸ and that, in turn, modifies the availability, absorption and accumulation in different plant parts.

Plant nutrients affect the activity and bioavailability of Cd in the soil–plant environment. Therefore, it is of great importance to study the interaction of Cd with plant nutrients in soil–plant systems to use it as a tool to minimize the accumulation of Cd in edible plant parts. The main objective of this review is to highlight the relationships of plant nutrients with Cd to understand how the management of plant nutrients is useful in reducing Cd availability in soil, root absorption, translocation to shoot and then grain, and how these nutrients are helpful in the development of tolerance against Cd toxicity and the reduction of Cd accumulation in plants, especially the edible parts.

FACTORS AFFECTING THE BIOAVAILABILITY OF CADMIUM

It is not necessary that all the Cd present in soil is available for plant uptake. The term bioavailability is defined as 'the part of the total concentration of a chemical that is available to receptor cells (plants, microorganisms, etc.)'. The part of soil Cd concentration available to plants is called bioavailable Cd. This bioavailable concentration is the main concern regarding its uptake and accumulation in plants rather than the total concentration of Cd in soil. So in any study, both the total concentration as well as the bioavailable concentration should be measured. A number of factors affect Cd bioavailability in soil, including soil pH, organic matter presence of other ions, root exudates, types and cultivars of crop plants, and plant age. ^{59,60} These factors influence the solubility of Cd compounds and the release of Cd into the soil solution or affect the ability of plants to take up Cd from soil. ⁶¹ Some have positive effects while others have negative effects on Cd bioavailability.

Among soil factors affecting Cd availability, soil pH is the most important.⁶² There is an indirect linear relationship between soil pH and Cd bioavailability, i.e. with a decrease in soil pH, metal uptake by plants increases.^{63,64} So, it may be possible that in



Cd-contaminated acidic soils availability is greater as compared to neutral and alkaline soils. Cadmium availability in such soils can be decreased by manipulating soil pH through the use of various amendments including liming⁴¹ and application of base-rich fertilizers. Application of some fertilizers, such as NH₄⁺ fertilizers including urea, ammonium sulfate and monoammonium phosphate (MAP), can enhance Cd availability by lowering pH. 10,50 Organic matter content is another important soil factor that affects Cd availability in soil as it is capable of retaining metal cations. Organic matter acts as a primary sorbent of heavy metals in organic forest soils.⁶⁵ Sauvé et al.⁶⁶ conducted a study in Canada on organic forest soils, and found that soil pH and total metal concentration were not consistent indicators of bioavailable Cd. The sorption affinity of soil for Cd was 30 times greater due to presence of organic matter as compared to mineral soil. So, in Cd-contaminated soils, use of various organic amendments such as farm yard manure (FYM), composts, bio-solids and bio-solid compost can effectively reduce Cd availability to plants. 40

The presence of other ions also has a great influence on Cd availability. This influence may be due to ionic strength,⁶⁷ complexation⁶⁸ and competition for soil exchange sites or root surface exchange sites.⁴¹ Ionic strength of the growth medium affects Cd availability inversely, which indicates that the lower the ionic strength of growth medium, the higher the metal concentration taken up by plants.⁶⁷ Cadmium is also found to form complexes in the presence of Cl⁻ which results in increasing solubility and thus availability of Cd.⁶⁸ Cations like Ca²⁺, Mg²⁺, Zn²⁺ and Mn²⁺ compete with Cd for uptake by plants^{41,42,69} and for exchange sites in soils.⁶⁸

Different plant species vary greatly in metal uptake from soil.⁷⁰ Some plants, including maize, pea, oat and wheat, are low accumulators of heavy metals, while some leafy vegetables, like spinach and lettuce, accumulate a high metal concentration in their leaves. 40 In general, Cd accumulation is greater in leaf vegetables, less in root vegetables and lowest in grain crops.⁷¹ Wheat accumulates much higher Cd concentrations than other cereal species. 40 The behavior of heavy-metal accumulation also varies from cultivar to cultivar as there are large genotypic differences for metal accumulation in many species.30-32 Jamali et al.32 conducted a pot culture experiment on four newly bred wheat cultivars (Anmol, TJ-83, Abadgar and Mehran-89) to reduce the potential of heavy-metal accumulation from amended sludge. The results showed that in two wheat cultivars (TJ-83 and Mehran-89) heavy-metal concentration was more than in the other two cultivars (Anmol and Abadgar) grown under similar amended sludge treatments.

The release of root exudates is an important plant factor that affects Cd bioavailability. Root exudates are part of the photosynthates of plants (12–40%) that are released through roots into soil, consisting of sugars, polysaccharides, amino acids, peptides, proteins and some organic acids.⁷² Organic acids released by plants can bind and sequester heavy metals in soil⁷³ and this mechanism protects roots from the toxic effects of heavy metals⁷⁴ and may also decrease Cd uptake by plants. As these exudates are photosynthates of plants, by improving plant nutrition, release of root exudates can be modified.

ESSENTIAL PLANT NUTRIENTS

Proper nutrition is a basic requirement of every living organism. There are now 17 elements which are considered essential for plants to complete their life cycle. These essential plant

nutrients are divided into two categories: macronutrients and micronutrients. Macronutrients include carbon (C), hydrogen (H), oxygen (O), nitrogen (N), phosphorus (P), potassium (K), calcium (Ca), magnesium (Mg) and sulfur (S). Micronutrients are zinc (Zn), copper (Cu), iron (Fe), manganese (Mn), boron (B), molybdenum (Mo), chlorine (Cl) and nickel (Ni).⁷⁵ Although silicon (Si) is not essential, it is considered a beneficial plant nutrient. These plants nutrients are not only required for better plant growth and development, but also helpful to alleviate different kinds of stresses like heavy-metal stress.^{41,47,56} The role of some important macronutrients and micronutrients, along with Si, as antagonists to Cd accumulation in grain/edible parts to minimize its dietary intake is discussed here.

Nitrogen

Nitrogen (N) is an essential macronutrient deficient in most soils, especially in arid and semi-arid regions. 76 It is an important component of many structural, genetic and metabolic compounds in plants, 22,77 and of the total nutrients absorbed by plant roots, 80% is contributed by N. 78 It is taken up by plants both in the form of ammonium (NH4 $^+$) and nitrate (NO3 $^-$), and in plants NO3 $^-$ is also reduced to NH4 $^+$ for assimilation into plant organic N. 56 For better crop growth, a combination of ammonium (NH4 $^+$) and nitrate (NO3 $^-$) sources is preferred. 79 Possible mechanisms to minimize Cd accumulation in cereals and legumes by improving N nutrition are presented in Fig. 1.

Rhizosphere chemistry, especially pH, is very important in determining the availability and absorption of nutrients and metals including Cd from soil. 80,81 Roots have a profound influence on soil pH and it has been found that, in general, pH of the rhizosphere is lower than bulk soil. Rhizosphere acidification occurs as a result of NH₄⁺ nutrition due to the release of protons (H⁺) by root cells or nitrification of NH₄+, and this induced acidification can promote mobilization of a localized metal in neutral to alkaline soil contaminated with a particular heavy metal like Cd.50,82 The type of N fertilizers applied will determine whether there would be a decrease or an increase in Cd uptake with its application. Compared to NO₃⁻ fertilizers, NH₄⁺-containing fertilizers could result in enhanced Cd uptake due to a decrease in soil pH. However, the effect of counter ions in fertilizers like Ca in Ca(NO₃)₂ cannot be overlooked. If applied at high rates, Ca in fertilizers could replace Cd present on soil particles, resulting in higher Cd concentration in the soil solution. However, this additive effect of counter ions like Ca on increased solubility and uptake of Cd is only possible at higher pH. At lower pH, the major effect on increased Cd uptake from soil would be of NH₄⁺ leading to a decrease in soil pH. In contrast to many researchers, Xie et al.83 recently investigated the effect of N form (NO_3^- vs NH_4^+) on growth and uptake of Cd and Zn by Thlaspi caerulesscens in hydroponics and rhizobox experiments and found that plants fed NO₃⁻ accumulated much more Cd than plants supplied with NH₄+, even though the rhizosphere pH was lower in plants treated with NH₄⁺. This NO₃⁻-induced Cd accumulation was speculated with the increase in organic acid exudation due to the accumulation of NO₃⁻ at the root surface. These contrasting results could also be related to the plant species used in the various experiments.

Contrasting results have been reported regarding the influence of N fertilizer on the uptake of Cd from soils. Increased N application would result in more biomass production by increasing the photosynthetic rate. As a result, Cd concentration in wheat grain would decrease due to the dilution effect⁵¹ as more biomass production sequesters more Cd in vegetative parts and very little



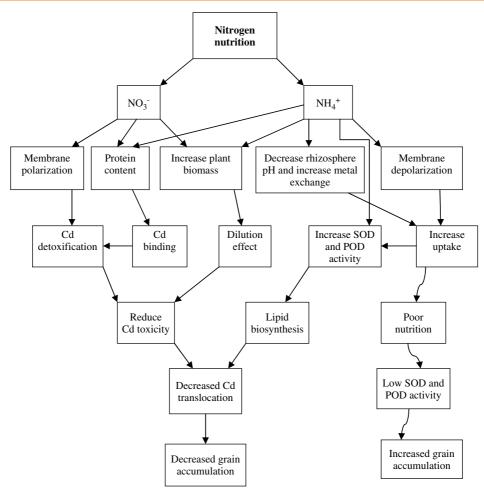


Figure 1. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving nitrogen nutrition (adapted from References 50–52,56,82).

moves into the grain.⁵¹ But Wangstrand *et al.*⁸⁴ claimed that with application of a higher rate of N, aimed not to enhance biomass production but to increase protein content of bread wheat grain, more Cd might be accumulated in plants. Time of application might be important, i.e. if an extra dose of N is applied at the vegetative stage, it can enhance biomass production; however, during the grain filling stage, the application of an extra dose of N can increase grain Cd concentration as NH₄⁺ fertilization increases the availability of metal ions due to soil acidification.⁵⁰ So, it is recommended that in Cd-contaminated soils more N application should be done at the vegetative stage to increase biomass production, while high N doses during the grain filling stage to increase protein content should be avoided. This strategy can reduce the Cd accumulation in grain.

One possible mechanism for alleviating Cd toxicity by the dilution effect might be the increase in soluble protein content⁵² which causes sequestration of the mobile form of Cd to the immobile form by binding to some protein molecules. The dilution of Cd concentration in the plant may also prevent membrane damage which is due to the high concentration of Cd on membrane surfaces. Pankovic *et al.*⁵² performed an experiment to analyze the effect of N nutrition on photosynthesis in Cd-treated sunflower plants. Three low levels of Cd (0.5, 2 and 5 mmol m⁻³) in combination with three N treatments (2, 7.5 and 10 mol m⁻³) were applied to young sunflower plants. They found that at 7.5 mol m⁻³

N, when soluble protein and Rubisco content were maximized, inhibition of photosynthetic activity by Cd was the lowest. From this study, it was concluded that N supply could be manipulated as a means of decreasing Cd toxicity to plants but optimum [N] to [Cd] ratios must be determined for specific plant species and growth conditions.⁵²

Ammonium ions cause cell membrane potential depolarization, which results in the influx of NH₄⁺ into the cytoplasm of the root cells.⁵⁰ This increased uptake of NH₄⁺ reduces the Cd uptake by cells. But this mechanism increased the translocation of Cd from root to shoot in sunflower plants possibly due to lack of a detoxification mechanism.⁵⁰ However, activities of superoxide dismutase (SOD) and peroxidase (POD) increased in the case of NH₄⁺ nutrition, which is considered a protective mechanism against stress. Peroxidase is also involved in lipids biosynthesis which acts as a physical barrier against heavy metals. 56 On the other hand, in plants fed with NO₃ most of the Cd accumulated in roots due to the detoxification mechanism.⁵⁰ Jalloh et al.⁵⁶ conducted a pot experiment to study the effects of various N fertilizer forms on antioxidant capacity and grain yield of rice (Oryza sativa L.) under Cd stress. They observed a higher Cd concentration and less N accumulation in plants treated with NO₃⁻-N, and the opposite (results) in the case of NH₄⁺-N treatment. Observation of plants indicated antagonistic interactions between NH₄⁺-N and Cd, and synergetic interactions between NO₃⁻-N and Cd. An increase in



SOD and POD activities was also more significant in plants treated with NH₄⁺-N, which is considered a protective mechanism against Cd stress. The authors argued that Cd stress could be alleviated by choosing a specific form of N fertilizer.

In light of the above findings, it can be concluded that crop species and the N source, rate and timing of application are important considerations in order to understand the role of N to minimize Cd accumulation in edible portions of crop plants. Still, further research is needed to fully understand how N nutrition plays its role to minimize Cd accumulation in plants, through mechanisms at the molecular level.

Phosphorus

The quantity of P in soils is less than N and K. Total P concentration in surface soils varies from 0.005 to 0.15%. 85 After N, it is the second most deficient plant nutrient and is applied to plants as fertilizers. More than 30 million metric tonnes of P_2O_5 in phosphate fertilizers per year are used worldwide, of which more than 99% is derived from rock phosphate. 86 The use of P fertilizers has increased crop production many fold, possibly through narrowing the N:P ratio. However, the use of phosphatic fertilizers can cause the contamination of soils with trace heavy metals like Cd, Cu, Mn, Ni, Pb and Zn which are naturally present in rock phosphate. 87,88 Possible mechanisms to minimize Cd accumulation in cereals and legumes by appropriate P nutrition are shown in Fig. 2.

Application of some P fertilizers like mono-ammonium phosphate (MAP) could increase solubility of Cd by lowering soil pH, resulting in enhanced accumulation of Cd from soils. In field experiments conducted in Manitoba, Canada, Grant and Bailey 10,89 studied the influence of P and Zn fertilizer management on Cd accumulation in flaxseed and durum wheat. They found that both concentration and accumulation of Cd in flaxseed and durum wheat increased by the application of P as MAP. In another study in Manitoba, application of MAP and potassium chloride (KCI) tended to increase Cd concentration in malting barley.90 These increased concentrations of Cd in grain might be due to increased solubility of Cd in soils which accelerated the Cd uptake by plants. The effect of P fertilizer application on the solubility of Cd and Zn in soils was investigated by Lambert et al.⁸⁸ In both field and laboratory experiments, they found that application of P fertilizers containing Cd and Zn enhanced Cd in the soil extract, while

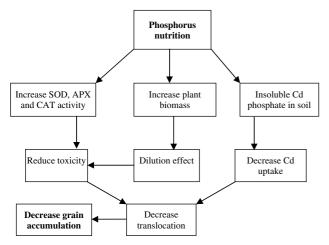


Figure 2. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving phosphorus nutrition (adapted from References 25,27,44,92–94,96,98).

Zn concentration in almost half of the treatments decreased as compared to the control. However, in neutral/alkaline soils, application of fertilizer MAP causes slight change in pH.⁹¹ This minute change in pH will increase availability of a localized heavy metal (Cd) to a very small extent.

Phosphate fertilizers on the other hand, also decrease the mobility of Cd in soil by changing mobile forms of Cd to an immobile form of Cd phosphate, 27,44 which is considered a key (extrinsic) mechanism of decreasing Cd availability to plants. Matusik et al.44 compared the effect of different P compounds (K₂HPO₄, NH₄H₂PO₄ and 'Polifoska 15' fertilizer) over a pH range of 4.0-9.0 and for reaction times of 2-1440 h. A reduction of greater than 99% in available Cd concentration was observed for all forms of phosphate between pH 6.75 and 9.00. Bolan et al. 92,93 explained phosphate-induced immobilization of Cd in soils by: (1) phosphate-induced Cd²⁺ adsorption; and (2) precipitation of Cd as Cd(OH)₂ and Cd₃(PO₄)₂. Several mechanisms were advanced for the immobilization mentioned which include: (1) an increase in pH; (2) an increase in surface charge; (3) co-adsorption of phosphate and Cd as ion pair; and (4) the surface complex formation of Cd on the phosphate compound.

Phosphorus application may increase plant growth and yields which can neutralize the toxic effect of Cd by the dilution effect. Wang et al.⁹⁴ found that addition of external P reduced toxic effects of Cd on P-treated plants as compared to the control (no P applied). In a greenhouse study, Dheri et al.²⁷ reported that application of P fertilizer as KH₂PO₄ to spinach (Spinacia oleracea L.) decreased the concentration of Cd in spinach plants by reducing the diethylene triamine pentaacetic acid (DTPA)-extractable Cd in soil, while also enhancing dry matter yield of both shoot and root. They concluded that application of P was effective for in situ immobilization of Cd in mildly Cd-contaminated soil, and their findings suggested that Cd toxicity in spinach plants grown in mildly Cd-contaminated soils can be minimized by phosphate-P application. Phosphorus is involved in glutathione (GSH) biosynthesis⁹⁵ which is commonly suggested as a precursor in phytochelatin (PC) synthesis. These phytochelatins compartmentalize Cd into vacuoles by forming Cd/PC complexes.⁴⁸ Wang et al.⁹⁴ observed an increase in GSH content in P-treated plants as compared to the zero-P control.

Reactive oxygen species (ROS) are continuously produced as byproducts of different metabolic reactions particularly under stress, causing damage to biomolecules like membrane lipids and proteins. Certain antioxidant enzymes are produced by plants in response to oxidative stress to mitigate the effect of ROS such as SOD, ascorbate peroxidase (APX) and catalase (CAT). So Superoxide dismutase converts the superoxide radical into hydrogen peroxide (H_2O_2) and oxygen (O_2). Hydrogen peroxide is still damaging and is further converted to H_2O by the action of CAT and APX. Wang *et al.* 4 found that the activity of these antioxidants increased under Cd stress by the application of P, which mitigates the oxidative stress and prevents membrane damage. So P can partly alleviate Cd toxicity. The actual mechanism is still unclear and needs further research.

In summary, there is no well-documented evidence to identify the intrinsic mechanism for the development of plant tolerance against Cd by P nutrition. Extensive research is required to understand the role of P nutrition in developing plant tolerance against Cd.

Sulfur

Sulfur (S) is an essential macronutrient that plays a key role in protein synthesis. It is an important structural component of many



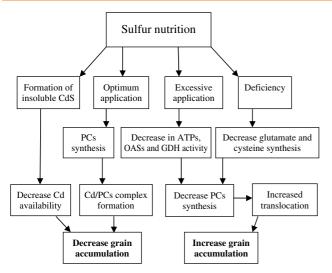


Figure 3. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving sulfur nutrition (adapted from References 48,49,72,104–106).

co-enzymes and prosthetic groups such as feredoxins important in N assimilation. Thus, S plays an important role in plant growth and development. Sulfur deficiency has become a recently emerging problem, which results in decreased crop yield and quality.⁹⁹ In the past, application of N fertilizers like ammonium sulfate and ammonium superphosphate, and atmospheric S depositions were enough to meet the S requirements of crops. However, at present, atmospheric S emissions have been reduced to a great extent and the use of high analysis fertilizers like urea and triple superphosphate or di-ammonium phosphate have caused S deficiency in many soils.¹⁰⁰ Possible mechanisms to reduce Cd accumulation in cereals and legumes by proper S nutrition are presented in Fig. 3.

Sulfur causes the production of H₂S in soil, which results in the formation of insoluble CdS making Cd unavailable to plants. This is due to the fact that application of sulfate fertilizers causes vulcanization in the soil and produces H₂S.⁷² Hassan et al.²² investigated the effect of different N fertilizer forms on Cd uptake and its toxicity in rice plants, using two Cd levels (0 and 1 μ mol) and three N forms (NH₄)₂SO₄, NH₄NO₃ and Ca(NO₃)₂. The minimum growth inhibition, and the highest photosynthetic rate and chlorophyll content were recorded in plants fed (NH₄)₂SO₄. Hassan et al.²² speculated that S in nutrient solution reduces the availability of Cd by forming insoluble CdS. On the other hand, Mclaughlin et al. 9 observed that under irrigation using high sulfate water content, the Cd uptake by crop plants was high due to formation of sulfato complexes. Sulfur also enhances glutathione (GSH) synthesis which is considered an important defensive mechanism against metal stress.

Cadmium toxicity can induce deficiency of several macronutrients in plants. ¹⁰¹ So, it is possible to minimize some of the Cd-induced negative effects through the optimum use of mineral nutrients. ⁸ Sub-optimal S nutrition in Cd-exposed plants can induce S deficiency due to synthesis of PC. ^{102,103} Metals like Cd are detoxified by chelation of metal ions with high-affinity phytochelatins, which seems an important defensive response against metal toxicity. ⁴⁹ But the availability of reduced S is an essential factor for PC synthesis. ¹⁰⁴ Cobbett ¹⁰⁵ proposed that PCs reduce activity of Cd in cytosol by forming Cd-PC complexes. These complexes are compartmentalized into vacuoles possibly by means of

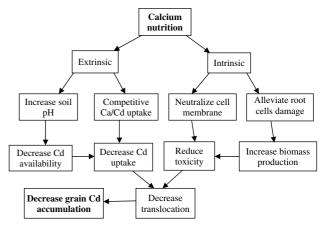


Figure 4. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving calcium nutrition (adapted from References 41.46.55.108).

an ATP-binding transporter present in tonoplast.⁴⁸ But the availability of reduced S is essential for PC synthesis.¹⁰⁴ So, optimum S nutrition is helpful for reducing Cd translocation within the plant body.

Cadmium toxicity induces rapid synthesis of PC thiol-based complexing substances. ¹⁰³ Phytochelatins are glutamate- and cysteine-rich proteins, so sulfate and nitrate assimilation pathways are important in PC synthesis: S metabolism enzymes are required for synthesis of cysteine, while enzymes of N metabolism are essential for synthesis of glutamate. ¹⁰⁶ In S-deficient Cd-treated plants, ATP sulfutylase (ATPS), *O*-acetylserine (OAS) and glutamate dehydrogenase (GDH) are increased, maybe partly due to a defensive mechanism, which is based on PC synthesis. ¹⁰⁶ Thus, optimum S nutrition should be recommended, which is not only important for better plant growth and development, but is also required for detoxification of Cd by PC synthesis, and to develop tolerance in plants against Cd toxicity.

Calcium

Calcium (Ca) is a macronutrient essential for plants as well as animals. It occupies much of the exchange sites in neutral and calcareous soils, while in acidic soils Ca²⁺ and Al³⁺ dominate the cation exchange capacity.⁸⁵ Plants absorb Ca²⁺ from the soil solution via mass flow and root interception. Possible mechanisms to check Cd accumulation in cereals and legumes by improving Ca nutrition are shown in Fig. 4.

In some studies, it has been shown that Ca as a plant nutrient can help to alleviate Cd toxicity. Both Ca and Cd compete for the same Ca channels in plants. ⁴⁶ Cadmium toxicity in animals may be affected by interactions between Ca and Cd. ¹⁰⁷ Choi et al. ¹⁰⁸ showed that plant Cd tolerance in tobacco plants was increased by Ca application due to formation of Cd – Ca-containing crystals, and removal of these crystals through the head cells of trichomes. Suzuki⁵⁵ investigated the effect of Cd toxicity on root growth inhibition in *Arabidopsis* seedlings supplemented with Ca. Retardation of root growth was observed, increasing cell death at growing points (root tips). On the other hand, application of Ca alleviates root growth inhibition due to a decrease in cell death.

Several mechanisms have been proposed to explain the effect of Ca on the alleviation of Cd toxicity. This may be due to the fact that the plasma membrane surface is usually negatively charged, and a high concentration of Ca²⁺ would neutralize the membrane



surface and minimize the toxic effect of Cd. It may also be due to the high concentration of ${\rm Ca^{2+}}$ around Ca channels which may decrease the influx of Cd.

Tlustos et al.⁴¹ determined the effect of liming with CaO and CaCO₃ on metal ion uptake by spring wheat in both pot and rhizobox experiments. They observed a 50% decrease in uptake for Cd, 80% for Zn and 20% for Pb. This can be explained on the basis of increased soil pH, which immobilizes metals in the soil. Another possible reason might be the high availability of Ca²⁺, which decreases the uptake of other metals. So, Ca can be helpful to alleviate Cd toxicity (at least partly) in the soil–plant environment.

Zinc

Zinc (Zn) is an important micronutrient essential for plant growth and development. Zinc status in soils and plants has a vital role in plant Cd accumulation. Previous research to understand Cd and Zn interactions has shown inconsistent results. Wu and Zhang ¹⁰⁹ proposed that the application of Zn can alleviate the physiological damage caused by Cd toxicity. In an hydroponic experiment, Hassan *et al.*⁵⁴ reported that application of Zn alleviated Cd toxicity by improving photosynthesis. Hart *et al.*⁴² suggested that Zn supplementation reduced Cd tissue concentration probably by inhibiting Cd uptake into roots. Possible mechanisms to prevent Cd accumulation in cereals and legumes by appropriate Zn nutrition are presented in Fig. 5.

Cadmium toxicity induces rapid synthesis of PC thiol-based complexing substances. ¹⁰³ Phytochelatins form Cd-PC complexes and Cd is sequestrated in the vacuole of root cells, resulting in decreased Cd translocation from roots to shoots. ⁵⁴ But in the presence of Zn, Zn-PC complexes are formed, ¹¹⁰ which can increase the concentration of free Cd and, as a result, Cd translocation from root to shoot is enhanced. Zhu *et al.* ¹¹¹ showed that Zn alleviated Cd toxicity in plants only at toxic Zn levels and did not improve plant growth, because in this case Zn toxicity suppressed plant growth. Another study showed that Cd-Zn interactions are synergistic to each other and increasing the concentrations of Cd and Zn in soils could enhance the

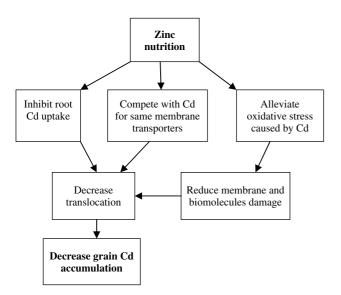


Figure 5. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving zinc nutrition (adapted from References 42,47,118,121).

accumulation of these metals in crops.¹¹² Adiloglu¹¹³ showed that Cd accumulation in plants is reduced with the application of Zn, and Cd accumulation is greater in Zn-deficient soils. Qiu *et al.*⁴⁷ proposed that increasing levels of Zn compete with Cd for the same membrane binding sites and transport systems and also limits Cd transport from phloem to grain. They showed that Zn had similar effects on Cd accumulation in winter wheat both in the long term (1 month) and the short term (24 h).

Chemical similarity between Zn and Cd is thought to be the main cause of Cd toxicity in higher plants due to their interactions with each other. Xue and Harrison¹¹⁴ identified a synergistic effect of Zn in relation to Cd uptake. They found that increasing the amount of Zn (>600 mg kg⁻¹) in soils containing high levels of Cd (10 mg kg⁻¹) resulted in a higher concentration of Cd in lettuce leaves. Similarly, Smilde *et al*.¹¹⁵ and Kachenko and Singh¹¹⁶ found high concentrations of Cd in leafy vegetables when soil Zn concentrations were increased. However, McKenna *et al*.¹¹⁷ identified a strong antagonistic effect of Zn on the accumulation of Cd in leafy vegetables at low Cd concentrations. Moreover, Chaney and Oliver⁸⁷ suggested that a Cd:Zn ratio of <1.5% in food effectively provides protection against Cd-induced health impacts.

Cadmium causes oxidation of NADPH, resulting in production of superoxide (O₂⁻),¹¹⁸ because Cd and other transition metals act as catalysts to induce rapid free radical synthesis (ROS). 119 Cakmak and Marschner¹²⁰ observed that production of O₂increases under Zn deficiency. So, it is possible that O_2 synthesis can be minimized by improving Zn nutrition. Aravind et al., 121 using Ceratophyllum demersum L. (a free-floating hydrophyte), investigated the protective effects of Zn against ROS induced by Cd. Different levels of Zn (10, 50, 100 and 200 μ mol Zn L⁻¹) against 10 μ mol Cd L⁻¹ were used. These authors observed that 10 μ mol Cd L⁻¹ triggered the production of H₂O₂ and O₂⁻, while addition of Zn (10-200 μ mol Zn L⁻¹) to Cd (10 μ mol Cd L⁻¹) significantly reduced the production of these reactive oxygen species (ROS). Aravind et al. 121 argued that Cd enhanced the oxidation of nicotine adenine dinucleotide phosphate (NADPH), resulting in O_2^- synthesis and the application of Zn inhibited NADPH oxidation and reduced O₂⁻ production. It is inferred that, by improving Zn nutrition, oxidative stress caused by Cd can be alleviated to a great extent.

Iron

Iron (Fe) is an essential micronutrient for both plants and animals. In neutral-to-alkaline soils, it is present as insoluble Fe³⁺ compounds. 122 However, under anaerobic conditions in soils, the ferrous (Fe²⁺) form is stable and readily available to plants. Plants reduce Fe³⁺ chelates to Fe²⁺ and transport via low-affinity iron transport systems through the plasma membrane, when Fe is not limiting. 123,124 Two distinct strategies are adopted by plants under Fe deficient conditions to assimilate Fe: (1) release of phyto-sidrophores (PS) by graminaceous monocotyledonous plants, which solubilize and take up Fe³⁺ by specific membrane receptors, and this Fe³⁺ is then reduced to Fe²⁺ in the cytoplasm (strategy II); and (2) acidification of the rhizosphere to enhance Fe availability, reduction of Fe³⁺ to Fe²⁺ by ferric reductase at the cell membrane and then transport to the cytoplasm (strategy I) in dicotyledonous and non-graminaceous monotyledonous plants. 123,124 Possible mechanisms to avoid Cd accumulation in cereals and legumes by proper Fe nutrition are presented in Fig. 6.

Ferrous ion (Fe²⁺) may compete with Cd for the same membrane transporters, so an efficient supply of Fe²⁺ to plants may decrease



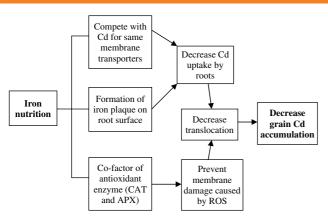


Figure 6. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving iron nutrition (adapted from References 125,126,129–131).

Cd uptake. But application of Fe^{2+} as $FeSO_4$ may oxidize to Fe^{3+} and become immobilized in soil. For this reason, Fe^{2+} should be applied to provide effective iron nutrition to plants. Sheng $et\ al.^{122}$ showed that soil application of $Fe\ fertilizer$ (EDTA– Na_2Fe) significantly reduced Cd concentration in rice grain, shoots and roots, while soil application of $FeSO_4$ and foliar application of both $FeSO_4$ and EDTA– Na_2Fe markedly enhanced the Cd concentrations in shoots and roots. From these results, it can be concluded that the type of $Fe\ fertilizer$ and the method of application might have marked effects on Cd accumulation in plants. Shao $et\ al.^{125}$ showed that Cd-induced oxidative stress in rice is alleviated by $Fe\ nutrition$.

Iron (Fe) is an integral cofactor of antioxidant enzymes such as catalase (CAT) and ascorbate peroxidase (APX). 126 So, by improving Fe nutrition, the activity of these enzymes may increase and this may prove an important defensive mechanism against ROS generated in the case of heavy metal stress. 96 However, free Fe is toxic to plants as it induces severe oxidative stress. 127 Sharma $et\,al.^{126}$ carried out a hydroponic experiment to investigate interference between Fe nutrition and Cd toxicity in barley seedlings. They observed increased CAT and APX activity under $25\,\mu\text{mol}\,\text{L}^{-1}$ Cd at all Fe levels except $250\,\mu\text{mol}\,\text{L}^{-1}$ where Fe induced CAT activity did not further increase due to Cd. The increased activity of antioxidant enzymes (CAT and APX) may alleviate the toxic effects of Cd by preventing membrane damage caused by ROSb. 25

Iron deficiency can induce accelerated Cd accumulation in plants by regulating soil pH and redox potential (E_h) , and this can be used as an important tool for phytoremediation of Cd from soil.¹²⁸ Formation of iron plaque on the surface of the root due to the release of O₂ and oxidants in the rhizosphere and the oxidation of Fe^{2+} to Fe^{3+} with the precipitation of iron oxide or hydroxide on the root surface is a common feature of aquatic plants such as rice. 129,130 Liu et al. 131 proposed that Fe plaque may adsorb and sequester Cd onto the root surfaces, and plant Fe nutrition can prevent Cd uptake by rice plants. Application of Fe to Cd-contaminated soils can increase Fe concentration and decrease Cd concentration in rice, leading to production of rice with high Fe but low Cd content, which is safe for humans. 131 Shao et al. 125 proposed that the inhibitory effect of high Fe nutrition on the expression of high affinity iron transport tends to reduce the uptake and accumulation of Cd in rice. They further hypothesized that both Fe and Cd compete for the common membrane bounded iron transport (carrier), so Cd toxicity in plants can be controlled by modifying Fe nutrition.

Manganese

Manganese (Mn) is a heavy metal as well as an essential micronutrient. It is also found to interact with Cd in soil–plant systems. Possible mechanisms to minimize Cd accumulation in cereals and legumes by adequate Mn nutrition are presented in Fig. 7. Availability of Mn to plants is partly decreased by Cd addition to soil. Ramachandran and D'Souza showed an antagonistic interaction between Cd and Mn, while Chen *et al.* indicated a synergetic interaction of Mn with Cd. Baszynski *et al.* suggested that Cd accumulation in plant tissues can be decreased by increasing the level of Mn possibly due to competition for the same membrane transporters.

Addition of Mn increases biomass production by improving root growth and enhancing $\mathrm{NO_3}^-$ uptake, 133 which reduces Cd toxicity. Alleviation of Cd toxicity prevents Cd translocation and accumulation in grain. 25 Cadmium replaces the central $\mathrm{Mg^{2+}}$ ion in the chlorophyll structure. 134 A high Mn level could alleviate Cd toxicity, especially by improving photosynthesis 53 because the application of Mn can partly restore the chloroplast structure damaged by Cd toxicity. 45

Silicon

Silicon (Si) is the second most abundant element in soil after oxygen. 135 It occurs in two major forms: silica and oxides of silicon, and both types exist in crystalline and/or amorphous forms such as quartz, flint, sandstone, opal and diatomaceous earth silicates. 136 In soil solution it occurs as silicic acid at concentrations ranging from 0.1 to 0.6 mmol L $^{-1}$, which is two orders of magnitude higher than macronutrient P. 137 Plants absorb most Si in mono-silicic acid form from the solution in the transpiration stream. Despite Si being a ubiquitous and prominent constitute of plants, it is still not widely recognized as an essential nutrient for plants. However, it has been proved to be beneficial for better plant growth and development, especially in plants of the *Gramineae* family. 43,138

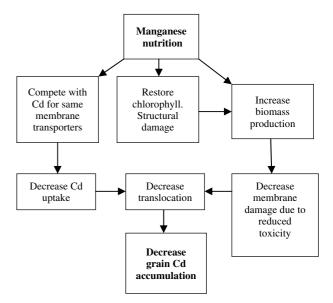


Figure 7. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving manganese nutrition (adapted from References 25,45,53,133,134).



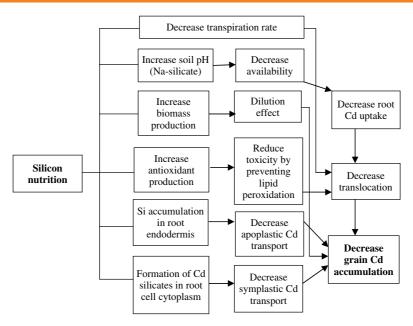


Figure 8. Possible mechanisms of minimizing Cd accumulation in cereals and legumes by improving silicon nutrition (adapted from References 135,140,143,147 – 149).

Possible mechanisms to reduce Cd accumulation in cereals and legumes by Si nutrition are presented in Fig. 8. Silicon can improve plant growth and tolerance to biotic and abiotic stresses. ^{137,139,140} In the case of heavy-metal stress, the presence of Si in the growth medium is helpful for reducing uptake and accumulation of heavy metals (like Cd) in rice, ⁴³ wheat ^{141,142} and maize ¹⁴³ seedlings.

Due to its high mobility in soil and plant system, Cd is readily taken up by plants. Among plant parts, roots are the major accumulator of Cd from their bathing medium. 144 The role of Si application in reducing Cd accumulation in edible plant parts has been well documented. 143,145,146 Increasing evidence is available showing that Si significantly interferes with root uptake and translocation of Cd from roots to shoots in plants. Based on a review of current literature, mechanisms responsible for low Cd accumulation in edible parts of plants are: (1) lower mobility of Si towards roots due to silicate induced pH rise in soils; (2) Si induced co-precipitation of Cd and Si in soil; (3) co-precipitation of Si and Cd at root surfaces; (4) decreased transport of Cd from roots to xylem; (5) reduced translocation of Cd from roots to shoots due to decreased evapotranspiration associated with Si deposition in cell walls, and as a double layer of polymerized Si in the cuticle; and (6) increased uptake of Ca with the application of Si, which decreased Cd uptake due to competition for uptake. The relationship between Si application and reduced Cd uptake has been extensively studied. As stated by Liang et al. 139 the mechanisms for Si-induced lower Cd in plants could be broadly classified as external factors (in soil) and internal factors (within the plant). In soils, application of Si as a chemical amendment resulted in significantly low phytoavailable Cd via increasing soil pH.¹⁴³ But this is not sufficient to explain the role of Si in the alleviation of Cd stress. Hodge 147 suggested that Si can change the plasticity of roots which is an important phenomenon in alleviating environmental stress. He proposed that the Si supply increased biomass production, which diluted the Cd concentration in shoots and roots. Moreover, application of Si increases Cd retention in roots and reduces Cd translocation to shoots by improving root growth.148

Cell wall bound Si in Si-accumulating plants inhibits Cd uptake significantly via the apoplastic pathway by covalently bonding with and capturing Cd as it diffuses through the cell wall and extra cellular spaces. 149 Silicon is capable of forming unstable silicates with heavy metals in the cytoplasm. This phenomenon inhibits symplastic transport of heavy metals. 140 In a review, Kirkham 63 concluded that few studies report Cd concentration in roots because of the difficulty in excavating them. Moreover, the author also concluded that it is also difficult to separate soil factors (e.g. an organic soil adhering to a root) that control Cd availability from root factors (e.g. root exudates). More studies were suggested by the author to distinguish Cd adsorption versus Cd absorption at the root, and what factors favor absorption over adsorption.

Foliar application of Si to reduce Cd accumulation in grain is a more effective and economical strategy, as compared to application of Si to the root. Liu *et al.*¹³⁵ studied the effect of foliar application of two silica sources on the alleviation of Cd stress and found that grain Cd accumulation in rice was significantly reduced by the application of Si, but had no significant effect on shoot Cd accumulation. The Cd–Si co-precipitation in cell walls restricts Cd translocation from shoot to grain, which alleviates the Cd toxicity and grain contamination.¹³⁵ The highest application efficiency of Si is obtained if it is applied at tillering to booting stage.¹⁵⁰ This may be due to the fact that plants absorb Si most effectively at this stage. Silicon nutrition can alleviate Cd toxicity to a great extent by improving plant tolerance to Cd stress. Concentration/rate of Si applied, placement method and timing of application (growth stage of the crop) are important considerations in this regard.

CONCLUSIONS

Cadmium is a toxic heavy metal for both plants and animals. In plants, Cd toxicity can cause decreases in photosynthetic rate, chlorophyll content, stomatal conductance, transpiration rate and relative leaf water content, and destruction of some physiological processes, which ultimately reduces plant growth and development. The consumption of toxic plant parts by

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animals and human beings can cause both acute and chronic disorders. Due to these risks, it is necessary to regulate Cd concentration in plants (primary producers) below permissible limits. The management of plant nutrients is very helpful to develop plant tolerance to Cd toxicity. Better plant nutrition can effectively alleviate Cd toxicity by a number of mechanisms. Application of some fertilizers, like phosphate fertilizers especially free from Cd, can decrease the availability of Cd either by fixation of Cd, or by forming insoluble or sparingly soluble phosphates of Cd. Appropriate application of plant nutrients increases plant biomass and grain yield, and decreases Cd concentration in grain and other edible plant parts by the dilution effect. Some plant nutrients, such as Ca, Fe and Zn, compete with Cd for the same membrane transporters. Plant nutrients also help to sequester Cd in vegetative parts by production of PCs and avoid Cd accumulation in grain. They help to alleviate physiological stress due to an excess of Cd. Thus, proper plant nutrition can be a good strategy to limit Cd in the food chain. The literature available, however, is insufficient to fully understand the role of plant nutrients to minimize Cd accumulation in grain or edible parts. Therefore, more research is required for a better understanding of the interactions between Cd and plant nutrients in soil-plant systems.

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REFERENCES

- 1 Chen F, Dong J, Wang F, Wu F, Zhang G, Li G, et al, Identification of barley genotypes with low grain Cd accumulation and its interaction with four microelements. Chemosphere 67:2082–2088 (2007).
- 2 Alloway BJ and Steinnes E, Anthropogenic additions of cadmium to soils, in *Cadmium in Soil and Plants*, ed. by Laughlin MJ and Singh BR. Kluwer Academic Publishers, Dordrecht, pp. 97–123 (1999).
- 3 di-Toppi LS and Gabrielli R, Response to cadmium in higher plants. Environ Exp Bot 41:105 – 130 (1999).
- 4 Wu FB, Chen FK, Wei KG and Zhang P, Effects of cadmium on free amino acids, glutathione, and ascorbic acid concentration in two barley genotypes (*Hordeum vulgare* L.) differing in cadmium tolerance. *Chemosphere* **57**:447–454 (2004).
- 5 Yang XE, Long XX, Ye HB, He ZL, Calvert DV and Stoffella PJ, Cadmium tolerance and hyperaccumulation in a new Zn-hyperaccumulating plant species (*Sedum alfredii* H.). *Plant Soil* **259**:181–189 (2004).
- 6 Jarup L, Hazards of heavy metal contamination. *Br Med Bull* **68**:167–182 (2003).
- 7 Nordberg GF, Cadmium and health in the 21st century historical remarks and trends for the future. *Biometals* **17**:485 489 (2004).
- 8 Hassan MJ, Zhu Z, Ahmad B and Mahmood Q, Infuence of cadmium toxicity on rice genotypes as affected by zinc sulfur and nitrogen fertilizers. *Caspian J Environ Sci* **4**:1–8 (2006).
- 9 McLaughlin MJ, Whatmuff M, Warne M, Heemsbergen D, Barry G, Bell M, et al, A field investigation of solubility and food chain accumulation of biosolid–cadmium across diverse soil types. Environ Chem 3:428–432 (2006).
- 10 Grant CA and Bailey LD, Effect of phosphorus and zinc fertilizer management on cadmium accumulation in flaxseed. J Sci Food Agric 73:307–314 (1997).
- 11 Ryan JA, Pahren HR and Lucas GB, Controlling cadmium in the human food chain: a review and rationale based on health effects. *Environ Res* **18**:251–302 (1982).
- 12 Nordic Council of Ministers, Cadmium Review. [Online] (2003). Available: http://www.who.int/ifcs/documents/forums/forum5/nmr_cadmium.pdf [accessed 10 January 2010].
- 13 Edwards R, Dixon DP and Walbot V, Plant glutathione S-transferases: enzymes with multiple functions in sickness and in health. *Trends Plant Sci* **5**:193–198 (2000).

- 14 Reeves PG and Chaney RL, Marginal nutritional status of zinc, iron, and calcium increases cadmium retention in the duodenum and other organs of rats fed a rice-based diet. *Environ Res* 96:311–322 (2004).
- 15 Chaney RL, Reeves PG, Ryan JA, Simmons RW, Welch RM and Angle JS, An improved understanding of soil Cd risk to humans and low cost methods to phytoextract Cd from contaminated soils to prevent soil Cd risks. *Biometals* 17:549–553 (2004).
- 16 Wagner GJ, Accumulation of cadmium in crop plants and its consequences to human health. *Adv Agron* **51**:173–212 (1993).
- 17 Codex Alimentarius Commission, Codex General Standard for Contaminants and Toxins in Foods. Report of the 34th Session of the Codex Committee on Food Additives and Contaminants, Rotterdam, The Netherlands. FAO/WHO, Rome, Italy, pp. 11–15 (2002).
- 18 Harris NS and Taylor GJ, Remobilization of cadmium in maturing shoots of near isogenic lines of durum wheat that differ in grain cadmium accumulation. *J Exp Bot* **52**:1473–1481 (2001).
- 19 European Commission, Commission regulation (EC) No 466/2001 of 8 March 2001. Setting maximum levels for certain contaminants in foodstuffs. Off J Eur Commun 77:1–13 (2001).
- 20 Popova LP, Maslenkova LT, Yordanova RY, Ivanova AP, Krantev AP and Szalai G, Exogenous treatment with salicylic acid attenuates cadmium toxicity in pea seedlings. *Plant Physiol Biochem* 47:224–231 (2009).
- 21 Xu J, Yin HX and Li X, Protective effects of proline against cadmium toxicity in micropropagated hyperaccumulator, Solanum nigrum L. Plant Cell Rep 28:325–333 (2009).
- 22 Hassan MJ, Wang F, Ali S and Zhang G, Toxic effects of cadmium on rice as affected by nitrogen fertilizer form. *Plant Soil* 277:359–365 (2005).
- 23 Chen Y and Huerta AJ, Effects of sulfur nutrition on photosynthesis in cadmium-treated barley seedlings. J Plant Nutr 20:845–856 (1997).
- 24 Sandalio LM, Dalruzo HC, Gomez M, Romero-Puetras MC and del-Rio LA, Cadmium-induced changes in the growth and oxidative metabolism of pea plants. J Exp Bot 52:2115–2126 (2001).
- 25 Ekmekçi Y, Tanyolaç D and Ayhan B, Effects of cadmium on antioxidant enzyme and photosynthetic activities in leaves of two maize cultivars. J Plant Physiol 165:600–611 (2008).
- 26 Verkleij JAC, Golan-Goldhirsh A, Antosiewisz DM, Schwitzguébel JP and Schröder P, Dualities in plant tolerance to pollutants and their uptake and translocation to the upper plant parts. *Environ Exp Bot* **67**:10–22 (2009).
- 27 Dheri GS, Brar MS and Malhi SS, Influence of phosphorus application on growth and cadmium uptake of spinach in two Cdcontaminated soils. J Plant Nutr Soil Sci 170:495–499 (2007).
- 28 Jackson AP and Alloway BJ, The transfer of cadmium from sewagesludge amended soils into the edible components of food crops. Water Air Soil Pollut 57 – 58:873 – 881 (1991).
- 29 Chaney RL, Ryan JA, Li YM and Brown SL, Soil cadmium as a threat to human health, in *Cadmium in Soils and Plants*, ed. by McLaughlin MJ, and Singh BR. Kluwer Academic Publishers, Dordrecht, pp. 219–256 (1999).
- 30 Yang YY, Jung JY, Song WY, Suh HS and Lee Y, Identification of rice varieties with high tolerance or sensitivity to lead and characterization of the mechanism of tolerance. *Plant Physiol* 124:1019–1026 (2000).
- 31 Liu J, Qian M, Cai G, Yang J and Zhub Q, Uptake and translocation of Cd in different rice cultivars and the relation with Cd accumulation in rice grain. *J Hazard Mater* **143**:443–447 (2007).
- 32 Jamali MK, Kazi TG, Arain MB, Afridi HI, Jalbani N, Kandhro GA, *et al*, Heavy metal accumulation in different varieties of wheat (*Triticum aestivum* L.) grown in soil amended with domestic sewage sludge. *J Hazard Mater* **164**:1386–1391 (2009).
- 33 Hinesly TD, Redborg KE, Ziegler EL and Alexander JD, Effect of soil exchange capacity on the uptake of Cd by corn. Soil Sci Soc Am J 46:490–497 (1982).
- 34 Florijin PJ and Van Beusiche ML, Cadmium distribution in maize inbred lines: Effects of pH and level of Cd supply. *Plant Soil* 153:79–84 (1993).
- 35 Grant CA, Clarke JM, Duguid S and Chaney RL, Selection and breeding of plant cultivars to minimize cadmium accumulation. Sci Total Environ 390:301 – 310 (2008).



- 36 Murakami M, Ae N and Ishikawa S, Phytoextraction of cadmium by rice (*Oryza sativa* L.), soybean (*Glycine max* (L.) Merr.), and maize (*Zea mays* L.). *Environ Pollut* 145:96–103 (2007).
- 37 Saifullah , Ghafoor A and Qadir M, Lead phytoextraction by wheat in response to the EDTA application method. *Int J Phytoremed* 11:268–282 (2008).
- 38 Saifullah, Meers E, Qadir M, de Caritat P, Tack FMG, Du Laing G, et al, EDTA-assisted Pb phytoextraction. Chemosphere 74:1279–1291 (2009).
- 39 Sas-Nowosielska A, Kucharski R, Malkowski E, Pogrzeba M, Kuperberg JM and Krynski K, Phytoextraction crop disposal – an unsolved problem. *Environ Pollut* 128:370–373 (2004).
- 40 Puschenreiter M, Horak O, Friesl W and Hartl W, Low-cost agricultural measures to reduce heavy metal transfer into the food chain – a review. *Plant Soil Environ* 51:1–11 (2005).
- 41 Tlustos P, Szakova J, Korinek K, Pavlikova D, Hanc A and Balik J, The effect of liming on cadmium, lead and zinc uptake reduction by spring wheat grown in contaminated soil. *Plant Soil Environ* 52:16–24 (2006).
- 42 Hart JJ, Welch RM, Norvell WA, Clarke JM and Kochian LV, Zinc effects on cadmium accumulation and partitioning in near isogenic lines of durum wheat that differ in grain cadmium concentration. *New Physiol* 167:391–401 (2005).
- 43 Shi XH, Zhang CH, Wang H and Zhang FS, Effect of Si on the distribution of Cd in rice seedlings. *Plant Soil* 272:53–60 (2005).
- 44 Matusik J, Bajda T and Manecki M, Immobilization of aqueous cadmium by addition of phosphates. J Hazard Mater 152:1332–1339 (2008).
- 45 Baszyński T, Wajda L, Król M, Wolińska D, Krupa Z and Tukendorf A, Photosynthetic activities of cadmium-treated tomato plants. Physiol Plant 48:365 – 370 (1980).
- 46 Nelson MT, Interactions of divalent cations with single calcium channels from rat brain synaptosomes. J Gen Physiol 87:201–222 (1986).
- 47 Qiu ZZ, Guan ZY and Long CY, Effect of zinc on cadmium uptake by spring wheat (*Triticum aestivum* L.): Long time hydroponic study and short time ¹⁰⁹Cd tracing study. *J Zhejjang Univ SCI* 6:643–648 (2005).
- 48 Salt DE and Rauser WE, MgATP-dependent transport of phytochelatins across the tonoplast of oat roots. *Plant Physiol* 107:1293 – 1301 (1995).
- 49 Hall JL, Cellular mechanisms for heavy metal detoxification and tolerance. J Exp Bot 53:1–11 (2002).
- 50 Zaccheo P, Laura C and Valeria DMP, Ammonium nutrition as a strategy for cadmium metabolisation in the rhizosphere of sunflower. Plant Soil 283:43–56 (2006).
- 51 Landberg T and Greger M, Influence of N and N Supplementation on Cd Accumulation in Wheat Grain, in 7th International Conference on the Biogeochemistry of Trace Elements, Uppsala '03, Conference Proceedings 1: III, Sweedish University of Agricultural Sciences, Uppsala, Sweden, pp. 90–91 (2003).
- 52 Pankovic D, Plesnicar M, Maksimoviic IA, Petrovic N, Sakac Z and Kastori R, Effect of nitrogen nutrition on photosynthesis in Cdtreated sunflower plants. Ann Bot 86:841 – 847 (2000).
- 53 Zornoza P, Vázquez S, Esteban E, Fernández-Pascual M and Carpena R, Cadmium-stress in nodulated white lupin: strategies to avoid toxicity. *Plant Physiol Biochem* 40:1003 – 1009 (2002).
- 54 Hassan MJ, Zhang G, Wu F, Wei K and Chen Z, Zinc alleviates growth inhibition and oxidative stress caused by cadmium in rice. J Plant Nutr Soil Sci 168:255 – 261 (2005).
- 55 Suzuki N, Alleviation by calcium of cadmium induced root growth inhibition in *Arabidopsis* seedlings. *Plant Biotechnol* 22:19–25 (2005).
- 56 Jalloh MA, Chen J, Zhen F and Zhang G, Effect of different N fertilizer forms on anti-oxidant capacity and grain yield of rice growing under Cd stress. J Hazard Mater 162:1081 – 1085 (2009).
- 57 Uri ND, Environmental considerations in the fertilizer use decision. Environ Geol 34:103–110 (1998).
- 58 Grant CA, Bailey LD, McLaughlin MJ and Singh BR, Management factors which influence cadmium concentrations in crops, in Cadmium in Soils and Plants, ed. by McLaughlin MJ Singh BR. Kluwer Academic Publishers, Dordrecht, pp. 151–158 (1999).
- 59 Harter RD and Naidu R, An assessment of environmental and solution parameter impact on trace-metal sorption by soil. Soil Sci Soc Am J 65:597 – 612 (2001).

- 60 Jung MC, Heavy metal concentration in soils and factors affecting metal uptake by plants in the vicinity of a Korean Cu–W mine. Sensors 8:2413–2423 (2008).
- 61 Mcbride MB, Reactions controlling heavy metal solubility in soils. Adv Soil Sci 10:1–56 (1989).
- 62 McBride M, Sauve S and Hendershot W, Solubility control of Cu, Zn, Cd and Pb in contaminated soils. Eur J Soil Sci 48:337 – 346 (1997).
- 63 Kirkham MB, Cadmium in plants on polluted soils: Effect of soil factors, hyperaccumulation and amendments. *Geoderma* **137**:19–32 (2006).
- 64 Kim KR, Owens G and Naidu R, Heavy metal distribution, bioaccessibility and phytoavailability in long-term contaminated soils from Lake Macquarie, Australia. Aust J Soil Res 47:166–176 (2009).
- 65 Ge Y and Hendershot W, Modeling sorption of Cd, Hg and Pb in soils by the NICA (non-ideal competitive adsorption) – Donnan model. Soil Sediment Contam 14:53 – 69 (2005).
- 66 Sauvé S, Manna S, Turmel MC, Roy AG and Courchesne F, Solid solution partitioning of Cd, Cu, Ni, Pb, and Zn in the organic horizons of a forest soil. *Environ Sci Technol* 37:5191–5196 (2003).
- 67 Göthberg A, Greger M, Holm K and Bengtsson BE, Influence of nutrient levels on uptake and effects of mercury, cadmium, and lead in water spinach. *J Environ Qual* 33:1247 – 1255 (2004).
- 68 Degryse, F, Buekers J and Smolders E, Radio-labile cadmium and zinc in soils as affected by pH and source of contamination. Eur J Soil Sci 55:113 – 121 (2004).
- 69 Ramachandran V and D'Souza TJ, Plant uptake of cadmium, zinc and manganese from four contrasting soils amended with Cd-enriched sewage sludge. J Environ Sci Health Toxic Hazard Subst Environ Eng 37:1337 – 1346 (2002).
- 70 Kloke A, Sauerbeck DR and Vetter H, The contamination of plants and soils with heavy metals and the transport of metals in terrestrial food chains, in *Changing Metal Cycles and Human Health*. Springer, Berlin, pp. 113–141 (1984).
- 71 Bingham FT, Bioavailability of Cd to food crops in relation to heavy metal content of sludge-amended soil. *Environ Health Perspect* 28:39–43 (1979).
- 72 Dong J, Mao WH, Zhang GP, Wu FB and Cai Y, Root excretion and plant tolerance to cadmium toxicity – a review. *Plant Soil Environ* 53:193 – 200 (2007).
- 73 Schwab AP, He YH and Banks MK, The influence of organic ligands on the retention of lead in soil. Chemosphere 61:856–866 (2005).
- 74 Liao M and Xie XM, Cadmium release in contaminated soils due to organic acids. *Pedosphere* 14:223–228 (2004).
- 75 Fageria NK, Filho MPB, Moreira A and Guimaraes CM, Foliar fertilization of crop plants. *J Plant Nutr* **32**:1044–1064 (2009).
- 76 Herandez LE, Garate A and Caroeba-Ruiz R, Effect of cadmium on the uptake, distribution and assimilation of nitrate in *Pisum sativum*. *Plant Soil* 189:97 – 106 (1997).
- 77 Tisdale SL and Nelson WL, Elements required in plant nutrition, in Soil Fertility and Fertilizers, ed. by Tisdale SL and Nelson WL. Macmillan, New York, pp. 66–104 (1975).
- 78 Marschner H, Mineral Nutrition of Higher Plants. 2. Academic Press, London, p. 889 (1995).
- 79 Frechilla S, Bonzalez EM, Royuela M, Arrese-Igor C, Lamsfus C and Aparicio-Tejo PM, Source of nitrogen nutrition affects pea growth involving changes in stomatal conductance and photorespiration. *J Plant Nutr* **22**:911 926 (1999).
- 80 Öborn I, Jansson G and Johnsson L, A field study on the influence of soil pH on trace element levels in spring wheat, potatoes and carrots. Water Air Soil Pollut 85:835–840 (1995).
- 81 Jansson G and Öborn I, Cadmium in Swedish carrots and the influence of soil factors. Acta Agric Scand Sect B Soil Plant 50:49 – 56 (2000).
- 82 Loosemore N, Straczek A, Hinsinger P and Jaillard B, Zinc mobilisation from a contaminated soil by three genotypes of tobacco as affected by soil and rhizosphere pH. *Plant Soil* 260:19–32 (2004).
- 83 Xie HL, Jiang RF, Zhang FS, McGrath SP and Zhao FJ, Effect of nitrogen form on the rhizosphere dynamics and uptake of cadmium and zinc by the hyperaccumulator *Thlaspi caerulescens*. *Plant Soil* 318:205–215 (2009).
- 84 Wangstrand H, Eriksso J and Oborn I, Cadmium concentration in winter wheat affected by nitrogen fertilization. *Eur J Agron* **26**:209–214 (2007).







- 85 Havlin JL, Tisdale SL, Nelson WL and Beaton JD, Soil Fertility and Fertilizer: An Introduction to Nutrient Management, 7th edition. Prentice Hall, NJ (2007).
- 86 International Fertilizer Industry Association (IFA), Fertilizers and Agriculture. [Online] (2005). Available: http://www.fertilizer.org/ifa/statistics/indicators/ind_products. asp Paris, France (2005) [accessed 10 January 2010].
- 87 Chaney RL and Oliver DP, Sources, potential adverse effects and remediation of agricultural soil contaminants, in *Contaminants and the Soil Environment in the Australasia–Pacific Region*, ed. by Naidu R, Kookana RS, Oliver DP, Rogers S and Mclaughlin MJ. Kluwer Academic, Dordrecht, pp. 323–359 (1996).
- 88 Lambert R, Grant C and Sauvé S, Cadmium and zinc in soil solution extracts following the application of phosphate fertilizers. *SciTotal Environ* **378**:293–305 (2007).
- 89 Grant CA and Bailey LD, Nitrogen, phosphorus and zinc management effects on grain yield and cadmium content in two cultivars of durum wheat. Can J Plant Sci 78:63-70 (1998).
- 90 Grant CA, Bailey LD and Therrien MC, Effect of N, P, and KCI fertilizers on grain yield and Cd concentration of malting barley. Fert Res 45:153–161 (1996).
- 91 Francois M, Grant C, Lambert R and Sauvé S, Prediction of cadmium and zinc concentration in wheat grain from soils affected by application of phosphate fertilizers varying in Cd concentration. *Nutr Cycl Agroecosyst* 83:125–133 (2009).
- 92 Bolan NS, Adriano DC, Mani S, Duraisamy P and Arulmozhiselvan S, Immobilization and phytoavailability of cadmium in variable charge soils: I. Effect of phosphate addition. *Plant Soil* 250:83–94 (2003).
- 93 Bolan NS, Duraisamy P and Mani S, Role of inorganic and organic soil amendments on immobilization and phytoavailability of heavy metals: a review involving specific case studies. *Aust J Soil Res* 41:1–23 (2003).
- 94 Wang H, Wang PF and Zhang H, Use of phosphorus to alleviate stress induced by cadmium and zinc in two submerged macrophytes. *Afr J Biotech* **8**:2176–2183 (2009).
- 95 May MJ, Vernoux T and Leaver C, Glutathione homeostasis in plants: implications for environmental sensing and plant development. J Exp Bot 49:649–667 (1998).
- 96 Gratão PL, Polle A, Lea PJ and Azevedo RA, Making the life of heavy metal stressed plants a little easier. Funct Plant Biol 32:32481–32494 (2005).
- 97 Davies MJ, Singlet oxygen-mediated damage to protein and its consequences. Biochem Biophys Res Commun 305:761 – 770 (2003).
- 98 Benavides MP, Gallego SM and Tomaro ML, Cadmium toxicity in plants. *Braz J Plant Physiol* **17**:21–34 (2005).
- 99 McGrath SP, Zhao FJ and Whiters PJA, Proceedings of the Fertilizers Society, in *The Fertilizers Society*. No. 379. Peterborough, Ontario (1996)
- 100 Tandon HLS, Sulphur Fertilizer for Indian Agriculture, A Guidebook, 2nd edition. Fertilizer Development and Consultation Organization, New Dehli, pp. 1–23 (1995).
- 101 Siedleska A, Some aspects of interactions between heavy metals and plant mineral nutrients. *Acta Soc Bot Pol* **64**:265 272 (1995).
- 102 McMahon PJ and Anderson JW, Preferential allocation of sulphur into γ -glutamylcysteinyl peptides in wheat plants grown at low sulphur in the presence of cadmium. *Physiol Plant* **104**:440–448 (1998).
- 103 Rauser WE, The role of thiols in plants under metal stress, in Sulphur Nutrition and Sulphur Assimilation in Higher Plants: Molecular, Biochemical and Physiological Aspects, ed. by Brunold C, Rennenberg H, De Kok LJ, Stulen I and Davidian JC. Haupt, Berne, pp. 169–183 (2000).
- 104 Rennenberg H, Brunold C, Dekok CJ and Stulen I, Sulphur Nutrition and Sulphur Assimilation in Higher Plants. SPB Academic Publishing, The Hague (1990).
- 105 Cobbett CS, Phytochelatins and their role in heavy metal detoxification. *Plant Physiol* **123**:825–833 (2000).
- 106 Astolfi S, Zuchi S and Passera C, Role of sulphur availability on cadmium-induced changes of nitrogen and sulphur metabolism in maize (Zea mays L.) leaves. J Plant Physiol 161:795–802 (2004).
- 107 Hinkle PM, Kinsella PA and Osterhoudt KC, Cadmium uptake and toxicity via voltage sensitive calcium channels. J Biol Chem 262:16333 – 16337 (1987).
- 108 Choi YE, Harada E, Wada M, Tsuboi H, Mortia Y, Kusano T, *et al*, Detoxification of cadmium in tobacco plant: formation and active

- excretion of crystals containing cadmium and calcium through trichomes. *Planta* **213**:45–50 (2001).
- 109 Wu FB and Zhang G, Genotypic differences in effect of Cd on growth and mineral concentrations in barley seedlings. Bull Environ Contam Toxicol 69:219–227 (2002).
- 110 Perrin DD and Watt AE, Complex formation in zinc and cadmium with glutathione. *Biochim Biophys Acta* **230**:96–104 (1971).
- 111 Zhu YG, Zhao ZQ, Li HY, Smith SE and Smith FA, Effect of zinc-cadmium interactions on the uptake of zinc and cadmium by winter wheat (*Triticum aestivum*) grown in pot culture. *Bull Environ Contam Toxicol* **71**:1289–1296 (2003).
- 112 Nan Z, Li J, Zhang J and Cheng G, Cadmium and zinc interactions and their transfer in soil crop system under actual field conditions. *Sci Total Environ* 285:187 – 195 (2002).
- 113 Adiloglu A, The effect of zinc (Zn) application on uptake of cadmium (Cd) in some cereal species. Arch Acker-Pfl Boden 48:553-556 (2002).
- 114 Xue Q and Harrison HC, Effect of soil zinc, pH and cultivar uptake in leaf lettuce (*Lactuca sativa* L. var. crispa). Commun Soil Sci Plant Anal 22:975–991 (1991).
- 115 Smilde B, Van Luit KW and Van Driel W, The extraction by soil and absorption by plants of applied zinc and cadmium. *Plant Soil* 143:233–238 (1992).
- 116 Kachenko AG and Singh B, Heavy metals contamination in vegetables grown in urban and metal smelter contaminated sites in Australia. Water Air Soil Pollut 169:101 – 123 (2006).
- 117 McKenna IM, Chaney RL and Williams FM, The effects of cadmium and zinc interactions on the accumulation and tissue distribution of zinc and cadmium in lettuce and spinach. *Environ Pollut* 79:113–120 (1993).
- 118 Kawano T, Kawano N, Muto S and Lapeyrie F, Cation-induced superoxide generation in tobacco cell suspension culture is dependent on ion valence. *Plant Cell Environ* **24**:1235 1241 (2001).
- 119 Ercal N, Gurer-Orhan H and Aykin-Burns N, Toxic metals and oxidative stress: Part I. Mechanisms involved in metal-induced oxidative damage. Curr Top Med Chem 1:529–539 (2001).
- 120 Cakmak I and Marschner H, Zinc-dependent changes in ESR signals, NADPH oxidase and plasma membrane permeability in cotton roots. *Physiol Plant* 73:182–186 (1988).
- 121 Aravind P, Prasad MNV, Malec P, Waloszek A and Strza3ka K, Zinc protects Ceratophyllum demersum L. (free-floating hydrophyte) against reactive oxygen species induced by cadmium. J Trace Elements Med Biol 23:50–60 (2009).
- 122 Sheng SG, Xue CM, Ying WD, Mei XC, Yun CZ and Fu ZX, Using iron fertilizer to control Cd accumulation in rice plants: A new promising technology. *Sci China C Life Sci* **51**:245 253 (2008).
- 123 Curie C and Briat JF, Iron transport and signaling in plants. *Ann Rev Plant Biol* **54**:183 206 (2003).
- 124 Schmidt W, Iron solutions: Acquisition strategies and signaling pathways in plants. *Trends Plant Sci* **8**:188–193 (2002).
- 125 Shao G, Chen M, Wang W, Mou R and Zhang G, Iron nutrition affects cadmium accumulation and toxicity in rice plants. *Plant Growth Regul* **53**:33–42 (2007).
- 126 Sharma SS, Kaul S, Metwally A, Goyal KC, Finkemeier I and Dietz KJ, Cadmium toxicity to barley (*Hordeum vulgare*) as affected by varying Fe nutritional status. *Plant Sci* 166:1287 – 1295 (2004).
- 127 Kampfenkel K, Montagu MV and Inze D, Effects of iron excess on Nicotiana plumbaginifolia plants. Implications to oxidative stress. Plant Physiol 107:725 – 735 (1995).
- 128 Bao T, Sun L, Sun T, Zhang P and Niu Z, Iron-deficiency induces cadmium uptake and accumulation in Solanum nigrum L. Bull Environ Contam Toxicol 82:338–342 (2009).
- 129 Chen CC, Dixon JB and Turner FT, Iron coatings on rice roots: morphology and models of development. *Soil Sci Soc Am J* **44**:1113–1119 (1980).
- 130 Taylor GJ, Crowder AA and Rodden R, Formation and morphology of an iron plaque on the roots of *Typha latifolia* L. grown in solution culture. *Am J Bot* **71**:666–675 (1984).
- 131 Liu H, Zhang J, Christie P and Zhang F, Influence of iron plaque on uptake and accumulation of Cd by rice (*Oryza sativa* L.) seedlings grown in soil. *Sci Total Environ* **394**:361–368 (2008).
- 132 Jarvis SC, Jones LHP and Hopper MJ, Cadmium uptake from solution by plants and its transport from roots to shoots. *Plant Soil* 44:179–191 (1976).



- 133 Balang PP, Kisová A, Pavlovkin J and Mistrík I, Effect of manganese on cadmium toxicity in maize seedlings. *Plant Soil Environ* 52:143–149 (2006).
- 134 Kupper H, Kupper F and Spiller M, In situ detection of heavy metal substitution chlorophylls in water plants. Photosynthesis Res 58:123–133 (1998).
- 135 Liu C, Li F, Luo C, Liu X, Wang S, Liu T, et al, Foliar application of two silica sols reduces cadmium accumulation in rice grains. J Hazard Mater 161:1466–1472 (2009).
- 136 Datnoff LE, Seebold KW and Correa-V FJ, The use of silicon for integrated disease management: reducing fungicide applications and enhancing host plant resistance, in *Silicon in Agriculture*, ed. by Datnoff LE, Snyder GH and Korndörfer GH. Elsevier, Amsterdam, pp. 171–184 (2001).
- 137 Epstein E, Silicon. Annu Rev Plant Physiol Plant Mol Biol 50:641–664 (1999).
- 138 Gong HJ, Zhu XY, Chen KM, Wang SM and Zhang CL, Silicon alleviates oxidative damage of wheat plants in pots under drought. *Plant Sci* 169:313–321 (2005).
- 139 Liang YC, Sun WC, Zhu YG and Christie P, Mechanisms of siliconmediated alleviation of abiotic stresses in higher plants: a review. *Environ Pollut* 147:422–428 (2007).
- 140 Neumann D and Niede U, Silicon and heavy metal tolerance of higher plants. Phytochemistry 56:685 – 692 (2001).
- 141 Cocker KM, Evans DE and Hodson MJ, The amelioration of aluminum toxicity by silicon in wheat (*Triticum aestivum* L.): malate exudation as evidence for an in plant mechanism. *Planta* 204:318 – 323 (1998).
- 142 Nowakowski W and Nowakowska J, Silicon and copper interaction in the growth of spring wheat seedlings. *Biol Plant* 39:463–466 (1997).

- 143 Liang YC, Wong JWC and Wei L, Silicon-mediated enhancement of cadmium tolerance in maize (*Zea mays L.*) grown in cadmium contaminated soil. *Chemosphere* 58:475–483 (2005).
- 144 Song A, Lib Z, Zhang J, Xue G, Fan F and Liang Y, Silicon-enhanced resistance to cadmium toxicity in *Brassica chinensis L*. is attributed to Si-suppressed cadmium uptake and transport and Si-enhanced antioxidant defense capacity. *J Hazard Mater* DOI:10.1016/j.jhazmat. 143 (2009).
- 145 Chen HM, Zheng CR, Tu C and Shen ZG, Chemical methods and phytoremediation of soil contaminated with heavy metals. Chemosphere 41:229–234 (2000).
- 146 Yu H, Wang JL, Fang W, Yuan JG and Yang ZY, Cadmium accumulation in different rice cultivars and screening for pollutionsafe cultivars of rice. *Sci Total Environ* **370**:302–309 (2006).
- 147 Hodge A, The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytol* **162**:9–24 (2004).
- 148 Zhang C, Wang L, Nie Q, Zhang W and Zhang F, Long term effects of exogenous silicon on cadmium translocation and toxicity in rice (Oryza sativa L.). Environ Exp Bot 62:300 – 307 (2008).
- 149 Wang L, Wang W, Chen Q, Cao W, Li M and Zhang F, Silicon-induced cadmium tolerance of rice seedlings. *J Plant Nutr* **23**:1397–1406 (2000)
- 150 Gan XQ, Jiang LG, Xu JY, Dong DF and Wei SQ, Characteristics and genotypic difference of silicon accumulation and distribution in rice. Plant Nutr Fert Sci 10:531 – 535 (2004).